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L1 19 FILE MEDLINE
L2 21 FILE CAPLUS
L3 13 FILE BIOSIS
L4 18 FILE EMBASE
L5 2 FILE WPIDS

TOTAL FOR ALL FILES

L6 73 CELL AND SCAFFOLD AND (LINEAR OR FUSION PARTNER OR TARGET?
SEQUE
NCE OR ADENOVIRUS)

=> s l6 and (dna or rna) and (polymerase or transcript? or nls or golgi or
mitochondria?)

L7 2 FILE MEDLINE
L8 3 FILE CAPLUS
L9 1 FILE BIOSIS
L10 1 FILE EMBASE
L11 0 FILE WPIDS

TOTAL FOR ALL FILES

L12 7 L6 AND (DNA OR RNA) AND (POLYMERASE OR TRANSCRIPT? OR NLS OR
GOLGI OR MITOCHONDRIA?)

=> dup rem l12

PROCESSING COMPLETED FOR L12

L13 4 DUP REM L12 (3 DUPLICATES REMOVED)

=> d 1-4 cbib abs

L13 ANSWER 1 OF 4 MEDLINE

1999199239 Document Number: 99199239. Factor-specific modulation of
CREB-binding protein acetyltransferase activity. Perissi V; Dasen J S;
Kurokawa R; Wang Z; Korzus E; Rose D W; Glass C K; Rosenfeld M G. (Howard
Hughes Medical Institute, University of California at San Diego, La
Jolla,

CA 92093-0648, USA.)PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF
THE UNITED STATES OF AMERICA, (1999 Mar 30) 96 (7) 3652-7. Journal code:
PV3. ISSN: 0027-8424. Pub. country: United States. Language: English.

AB CREB-binding proteins (CBP) and p300 are essential **transcriptional**
coactivators for a large number of regulated DNA-binding
transcription factors, including CREB, nuclear receptors, and
STATs. CBP and p300 function in part by mediating the assembly of
multiprotein complexes that contain additional cofactors such as p300/CBP
interacting protein (p/CIP), a member of the p160/SRC family of
coactivators, and the p300/CBP associated factor p/CAF. In addition to
serving as molecular **scaffolds**, CBP and p300 each possess
intrinsic acetyltransferase activities that are required for their
function as coactivators. Here we report that the **adenovirus** E1A
protein inhibits the acetyltransferase activity of CBP on binding to the
C/H3 domain, whereas binding of CREB, or a CREB/E1A fusion protein to the
KIX domain, fails to inhibit CBP acetyltransferase activity.

Surprisingly,

p/CIP can either inhibit or stimulate CBP acetyltransferase activity
depending on the specific substrate evaluated and the functional domains
present in the p/CIP protein. While the CBP interaction domain of p/CIP

inhibits acetylation of histones H3, H4, or high mobility group by CBP,
it enhances acetylation of other substrates, such as Pit-1. These
observations suggest that the acetyltransferase activities of CBP/p300
and p/CAF can be differentially modulated by factors binding to distinct
regions of CBP/p300. Because these interactions are likely to result in
differential effects on the coactivator functions of CBP/p300 for
different classes of **transcription** factors, regulation of
CBP/p300 acetyltransferase activity may represent a mechanism for
integration of diverse signaling pathways.

L13 ANSWER 2 OF 4 CAPLUS COPYRIGHT 1999 ACS

1996:58252 Document No. 124:78726 **DNA** construct for effecting
homologous recombination and uses for recombinant protein production.
Trecu, Douglas A.; Heartlein, Michael W.; Selden, Richard F.
(Transkaryotic Therapies, Inc., USA). PCT Int. Appl. WO 9531560 A1
19951123, 147 pp. DESIGNATED STATES: W: AM, AT, AU, BB, BG, BR, BY, CA,
CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ,
LK, LR, LT, LU, LV, MD, MG, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE,
SG, SI, SK, TJ, TM, TT; RW: AT, BE, BF, BJ, CF, CG, CH, CI, CM, DE, DK,
ES, FR, GA, GB, GR, IE, IT, LU, MC, ML, MR, NE, NL, PT, SE, SN, TD, TG.
(English). CODEN: PIXXD2. APPLICATION: WO 95-US6045 19950511.

PRIORITY:

US 94-243391 19940513.

AB The invention relates to constructs comprising: a) a **targeting**
sequence; b) a regulatory sequence; c) an exon; and d) an unpaired
splice-donor site. The invention further relates to a method of
producing
protein in vitro or in vivo comprising the homologous recombination of a
construct as described above within the **cell**. The homologously
recombinant **cell** is then maintained under conditions which will
permit **transcription** and translation, resulting in protein
expression. The present invention further relates to homologously
recombinant **cells**, including primary, secondary, or immortalized
vertebrate **cells**, methods of making the **cells**, methods
of homologous recombination to produce fusion genes, methods of altering
gene expression in the **cells**, and methods of making a protein in
a **cell** employing the constructs of the invention.

L13 ANSWER 3 OF 4 CAPLUS COPYRIGHT 1999 ACS

1994:48303 Document No. 120:48303 Binding specificity of a nuclear
scaffold: supercoiled, single-stranded, and **scaffold**
-attached-region **DNA**. Kay, Volker; Bode, Juergen (Ges.
Biotechnol. Forsch. m.b.H., Braunschweig-Stoeckheim, D-38124, Germany).
Biochemistry, 33(1), 367-74 (English) 1994. CODEN: BICHAW. ISSN:
0006-2960.

AB **Scaffold**-attached-region (SAR) elements of **DNA** enhance
transcriptional rates, and this has been correlated with their
ability to undergo sepn. into single strands (ssDNA) under conditions of
neg. superhelicity (Bode et al., 1992). The competition studies
presented
here suggest that the SAR-**scaffold** interaction is based, in
part, on the recognition of single strands, while about one-half of SAR
sites are inaccessible to ssDNA. Conversely, since there are 20,000 SAR
sites but more than 60,000 sites for ssDNA per nuclear equiv., not all
ssDNA sites are open for SARs. In addn., a completely sep. set of
binding
centers recognizing and enzymically converting **DNA** of
superhelical d. below -0.04 can be titrated. These findings reflect
multiple binding specificities for **scaffold** preps. that are

routinely used for screening **scaffold**-attached regions.

L13 ANSWER 4 OF 4 MEDLINE

DUPLICATE 1

95112899 Document Number: 95112899. An actin infrastructure is associated with eukaryotic chromosomes: structural and functional significance.

Sauman I; Berry S J. (Wesleyan University, Biology Department, Middletown,

CT 06459-0170..)EUROPEAN JOURNAL OF CELL BIOLOGY, (1994 Aug) 64 (2) 348-56. Journal code: EM7. ISSN: 0171-9335. Pub. country: GERMANY: Germany, Federal Republic of. Language: English.

AB The presence of actin in eukaryotic nuclei, and, especially, its functional significance has not been well established. We have found that under routine immunocytochemical conditions, no actin can be detected in insect follicle **cell** nuclei by means of antibody (both mono- and polyclonal) or phalloidin staining. However, a pretreatment of nuclear preparations with two different endonucleases (deoxyribonuclease I or micrococcal nuclease) to remove a substantial amount of chromosomal **DNA** uncovers the presence of nuclear actin for both antibody and phalloidin detection. Employing the same nuclease digestion followed by antibody or phalloidin staining with squash preparations of *Drosophila* polytene chromosomes revealed that the nuclear actin is directly associated with the chromosomes. A strong positive signal in the polytene chromosomes obtained with phalloidin labeling not only confirmed the presence of actin in the chromosomes, but indicates that a considerable amount of nuclear actin is present in filamentous form (F-actin) rather than monomeric (G-actin). The detection of actin associated with *Xenopus* embryo chromosomes suggests the significance of chromosomal actin for diploid vertebrate **cells**. Using the specific actin disrupting agent cytochalasin D, we have demonstrated the structural significance of nuclear actin in maintaining the **linear** integrity of polytene chromosomes. Further, we present evidence that **RNA polymerase** II closely interacts with the chromosomal actin **scaffold**, and that its association with chromosomes does not require the presence of **DNA**.

=> s nolan g?/au,in;s payan d?/au,in

'IN' IS NOT A VALID FIELD CODE

L14 89 FILE MEDLINE

L15 89 FILE CAPLUS

L16 147 FILE BIOSIS

'IN' IS NOT A VALID FIELD CODE

L17 82 FILE EMBASE

L18 10 FILE WPIDS

TOTAL FOR ALL FILES

L19 417 NOLAN G?/AU,IN

'IN' IS NOT A VALID FIELD CODE

L20 109 FILE MEDLINE

L21 99 FILE CAPLUS

L22 152 FILE BIOSIS

'IN' IS NOT A VALID FIELD CODE

L23 108 FILE EMBASE

L24 1 FILE WPIDS

TOTAL FOR ALL FILES

L25 469 PAYAN D?/AU,IN

=> s 119 and 125

L26 0 FILE MEDLINE
L27 1 FILE CAPLUS
L28 0 FILE BIOSIS
L29 0 FILE EMBASE
L30 1 FILE WPIDS

TOTAL FOR ALL FILES

L31 2 L19 AND L25

=> dup rem l31

PROCESSING COMPLETED FOR L31

L32 1 DUP REM L31 (1 DUPLICATE REMOVED)

=> d cbib abs

L32 ANSWER 1 OF 1 CAPLUS COPYRIGHT 1999 ACS DUPLICATE 1
1999:8108 Document No. 130:62960 Combinatorial enzymic complexes for drug
screening. **Payan, Donald**; Nolan, Garry P. (Rigel
Pharmaceuticals, Inc., USA). PCT Int. Appl. WO 9856904 A1 19981217, 71
pp. DESIGNATED STATES: W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA,
CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, GW, HU, ID, IL, IS,
JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW,
MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA,
UG, UZ, VN, YU, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM; RW: AT, BE, BF,
BJ, CF, CG, CH, CI, CM, CY, DE, DK, ES, FI, FR, GA, GB, GR, IE, IT, LU,
MC, ML, MR, NE, NL, PT, SE, SN, TD, TG. (English). CODEN: PIXXD2.
APPLICATION: WO 98-US11926 19980610. PRIORITY: US 97-873601 19970612.

AB The invention relates to the formation of novel in vivo combinatorial
enzyme complexes for use in screening candidate drug agents for
bioactivity. Thus, compns. of novel mixts. of enzymes in a spatially
constricted or defined manner, i.e., by binding of the enzymes to a
scaffold mol., are provided. This configuration of enzymes allows them
to
act on a precursor mol. in a novel or efficient manner to form a
candidate
bioactive agent which may then be screened for bioactivity. These
scaffolds, and the corresponding enzymes, are introduced into a variety
of
different types of cells, generally using retroviral introduction of the
nucleic acids encoding them. Precursor mols. are then added and the
cells
are screened for desired phenotypes.

=> s scaffold and (dna or rna) and (bind? or polymerase or transcript?) and
(target? sequence or nls or golgi or mitochondria?)

L33 3 FILE MEDLINE
L34 3 FILE CAPLUS
L35 2 FILE BIOSIS
L36 2 FILE EMBASE
L37 0 FILE WPIDS

TOTAL FOR ALL FILES

L38 10 SCAFFOLD AND (DNA OR RNA) AND (BIND? OR POLYMERASE OR
TRANSCRIPT

?) AND (TARGET? SEQUENCE OR NLS OR GOLGI OR MITOCHONDRIA?)

=> s l38 not (l32 or l12)

L39 0 S L32
 L40 3 FILE MEDLINE
 L41 1 S L32
 L42 2 FILE CAPLUS
 L43 0 S L32
 L44 2 FILE BIOSIS
 L45 0 S L32
 L46 2 FILE EMBASE
 L47 0 S L32
 L48 0 FILE WPIDS

TOTAL FOR ALL FILES

L49 9 L38 NOT (L32 OR L12)

=> s l38 not (l31 or l12)

L50 3 FILE MEDLINE
 L51 2 FILE CAPLUS
 L52 2 FILE BIOSIS
 L53 2 FILE EMBASE
 L54 0 FILE WPIDS

TOTAL FOR ALL FILES

L55 9 L38 NOT (L31 OR L12)

=> dup rem l 55

ENTER REMOVE, IDENTIFY, ONLY, OR (?):end

=> dup rem l55

PROCESSING COMPLETED FOR L55

L56 4 DUP REM L55 (5 DUPLICATES REMOVED)

=> d cbib abs 1-4

L56 ANSWER 1 OF 4 MEDLINE

1999138982 Document Number: 99138982. Open reading frame 1a-encoded subunits

of the arterivirus replicase induce endoplasmic reticulum-derived double-membrane vesicles which carry the viral replication complex. Pedersen K W; van der Meer Y; Roos N; Snijder E J. (Division of Electron Microscopy, Department of Biology, University of Oslo, Oslo, Norway.)JOURNAL OF VIROLOGY, (1999 Mar) 73 (3) 2016-26. Journal code: KCV.

ISSN:

0022-538X. Pub. country: United States. Language: English.

AB The replicase of equine arteritis virus (EAV; family Arteriviridae, order Nidovirales) is expressed in the form of two polyproteins (the open reading frame 1a [ORF1a] and ORF1ab proteins). Three viral proteases cleave these precursors into 12 nonstructural proteins, which direct both genome replication and subgenomic mRNA **transcription**.

Immunofluorescence assays showed that most EAV replicase subunits localize

to membranes in the perinuclear region of the infected cell. Using replicase-specific antibodies and cryoimmunoelectron microscopy, unusual double-membrane vesicles (DMVs) were identified as the probable site of EAV RNA synthesis. These DMVs were previously observed in cells infected with different arteriviruses but were never implicated in viral

RNA synthesis. Extensive electron microscopic analysis showed that they appear to be derived from paired endoplasmic reticulum membranes and that they are most likely formed by protrusion and detachment of vesicular

structures with a double membrane. Interestingly, very similar membrane rearrangements were observed upon expression of ORFla-encoded replicase subunits nsp2 to nsp7 from an alphavirus-based expression vector. Apparently, the formation of a membrane-bound scaffold for the replication complex is a distinct step in the arterivirus life cycle, which is directed by the ORFla protein and does not depend on other viral proteins and/or EAV-specific RNA synthesis.

L56 ANSWER 2 OF 4 CAPLUS COPYRIGHT 1999 ACS DUPLICATE 1

1998:768524 Document No. 130:148337 Molecular Basis for the DNA Sequence Selectivity of Ecteinascidin 736 and 743: Evidence for the Dominant Role of Direct Readout via Hydrogen Bonding. Seaman, Frederick C.; Hurley, Laurence H. (Drug Dynamics Institute College of Pharmacy, The University of Texas at Austin, Austin, TX, 78712, USA). J. Am. Chem. Soc., 120(50), 13028-13041 (English) 1998. CODEN: JACSAT. ISSN: 0002-7863. Publisher: American Chemical Society.

AB The marine natural product ecteinascidin 743 (Et 743) is currently in phase II clin. trials. The authors have undertaken parallel structural and modeling studies of an Et 743-(N2-guanine) 12-mer DNA adduct and an adduct involving the structurally related Et 736 of the same sequence to ascertain the structural basis for the ecteinascidin-DNA sequence selectivity. In contrast to the C-subunit differences found in Et 736 and Et 743, they have identical A-B-subunit scaffolds, which are the principal sites of interaction with DNA bases. These identical scaffolds generate parallel networks of drug-DNA hydrogen bonds that assoc. the drugs with the three base pairs at the recognition site. The authors propose that these parallel hydrogen bonding networks stabilize the Et 736 and Et 743 A- and B-subunit prealkylation binding complex with the three base pairs and are the major factors governing sequence recognition and reactivity. The possibility that a unique hydrogen-bonding network directs the course of sequence recognition was examd. by first characterizing the hydrogen-bonding substituents using 1H NMR properties of the exchangeable protons attached to the hydrogen-bond donor and other protons near the proposed acceptor. Using these exptl. findings as indicators of hydrogen bonding, Et 736-12-mer duplex adduct models (binding and covalent forms) contg. the favored sequences 5'-AGC and 5'-CGG were examd. by mol. dynamics (MD) to evaluate the stability of the hydrogen bonds in the resulting conformations. The MD-generated models of these favored sequences display optimal donor/acceptor

positions

for maximizing the no. of drug-DNA hydrogen bonds prior to covalent reaction. The results of MD anal. of the carbinolamine (binding) forms of the sequences 5'-GGG (moderately reactive) and 5'-AGT (poorly reactive) suggested reasons for their diminished hydrogen-bonding capability. These exptl. and modeling results provide the structural basis for the following sequence specificity rules: For

the

target sequence 5'-XGY, the favored base to the 3'-side, Y, is either G or C. When Y is G, then a pyrimidine base (T or C) is favored for X. When Y is C, a purine (A or G) is favored for X.

L56 ANSWER 3 OF 4 MEDLINE

95352643 Document Number: 95352643. Nuclear multicatalytic proteinase alpha subunit RRC3: differential size, tyrosine phosphorylation, and susceptibility to antisense oligonucleotide treatment. Benedict C M; Ren L; Clawson G A. (Department of Pathology, Pennsylvania State University,

Hershey 17033, USA.)BIOCHEMISTRY, (1995 Jul 25) 34 (29) 9587-98.

Journal
code: AOG. ISSN: 0006-2960. Pub. country: United States. Language: English.

AB Multicatalytic proteinases (MCPs) are macromolecular structures involved in intracellular degradation of many types of proteins. MCPs are composed of a 20S "core" which consists of both structural (alpha) and presumed catalytic (beta) subunits in association with complexes of accessory proteins. Immunohistochemical studies have shown MCP subunits to be largely cytoplasmic, although nuclear localization is also observed. Reverse **transcription/polymerase** chain reaction amplifications were performed with redundant primers to conserved regions within known subunits, in an attempt both to identify potential new subunits and to define the repertoire of subunits expressed in hepatocytes. No new subunits were identified, and we found that RRC3, an alpha subunit of MCPs which contains a putative nuclear localization signal (**NLS**), was the predominant alpha subunit expressed in hepatocytes and hepatocyte-derived cell lines. Antibodies were developed against a unique C-terminal peptide region of RRC3. Immunohistochemical studies using affinity-purified antibodies showed that RRC3 has both cytoplasmic and nuclear localizations. Immunoprecipitation/immunoblot analyses showed that a significant proportion of nuclear RRC3 was associated with the nuclear **scaffold** (NS). NS RRC3 showed a significantly smaller M(r) (24,000) than the cytoplasmic form (M(r) 28,000), and only the nuclear form contained phosphotyrosine. In metabolic labeling experiments with [32P]orthophosphate, the major nuclear and NS form observed showed an M(r) of 24,000, whereas no labeling of cytosolic RRC3 was observed. A minor 32P-labeled band of M(r) 28,000 was also observed in nuclei, and this M(r) 28,000 form was found in the soluble nuclear extract within MCP complexes. These results suggest that tyrosine phosphorylation of the cytosolic form (M(r) 28,000) rapidly triggers nuclear import, which is in turn quickly followed by conversion to the major M(r) 24,000 form associated with NS. Treatment with antisense oligonucleotides targeted to the initiation site of RRC3 reduced the growth of a hepatocyte-derived cell line by 95% and produced a marked morphological change (in the absence of overt toxicity). Under these treatment conditions, RRC3 mRNA was dramatically reduced. RRC3 protein was also dramatically reduced in the NS, but showed only a small reduction in cytosol, suggesting that the nuclear RRC3 may be important in cell growth and differentiation.

L56 ANSWER 4 OF 4 MEDLINE DUPLICATE 2
94187728 Document Number: 94187728. Molecular genetic analyses of a 376-kilodalton **Golgi** complex membrane protein (giantin) [retracted in Mol Cell Biol 1995 Jan;15(1):591]. Seelig H P; Schranz P; Schroter H; Wiemann C; Griffiths G; Renz M. (Institute of Immunology and Molecular Genetics, Karlsruhe, Germany.)MOLECULAR AND CELLULAR BIOLOGY, (1994 Apr) 14 (4) 2564-76. Journal code: NGY. ISSN: 0270-7306. Pub. country: United States. Language: English.

AB Molecular genetic analyses of a 376-kDa **Golgi** complex (GC) membrane protein (giantin) are described. The immunoglobulin G fraction of a human serum containing antibodies against GC antigens as revealed by indirect immunofluorescence microscopy with Hep-2 cells was used to screen a HeLa cDNA expression library, yielding four overlapping cross-hybridizing clones. Additional cDNA clones were retrieved from a lambda gt11 human thyroid cDNA library or generated by reverse **transcriptase**-mediated PCR from HeLa cell mRNA. Alignment of the

clones resulted in a consensus cDNA of 10,300 bp encoding a protein of

kDa. The corresponding mRNA with a size of about 10 kb was detected by Northern (RNA) blotting of HeLa, Hep-G2, and Jurkat cell RNA. Sequence analyses of the protein revealed an extraordinarily high content of heptad repeats with the probability of forming coiled coils similar to the proteins of the myosin family. Five overlapping recombinant proteins covering the entire sequence were synthesized and used for antibody production in rabbits and for affinity purification of human and rabbit antibodies. Indirect immunofluorescence experiments also done with brefeldin A-treated Hep-2 and Pt K1 cells revealed an identical GC staining of both the affinity-purified human and rabbit antibodies. Double labeling experiments with antibodies against the GC marker mannosidase II as well as immunoelectron microscopic studies confirmed

the

localization of the protein within the GC. A corresponding endogenous large-molecular-mass protein of about 390 kDa was found in [35S]methionine-labeled Hep-2 cell lysates as well as in GC-enriched subcellular fractions from rat liver. The protein as well as the recently described proteins golgin-95 and golgin-160 (M. J. Fritzler, J. C. Hamel, R. L. Ochs, and E. K. L. Chan, J. Exp. Med. 178:49-62, 1993) may belong

to

a new group of Golgi proteins with a high content of heptad repeats which may exert functions in scaffold formation or vesicle transport. As far as can be concluded from immunological and personally communicated partial cDNA sequence data, the protein seems to be identical with a 400-kDa Golgi protein (giantin) recently described (A. D. Linstedt and H. P. Hauri, Mol. Biol. Cell 4:679-693, 1993). Therefore, we agreed to adopt the name giantin.

=> dis his

Insert the Sequence Listing as pages 58-68.

Renumbe the Claims as pages 69-72.

Renumbe the Abstract as page 73.

In the Claims:

1. (Amended) A cell containing a composition comprising:

a) an [exogeneous]exogenous scaffold having no enzymatic activity and comprising at least a first binding site and a second binding site; and

b) at least a first and a second enzyme, wherein at least one of said enzymes is heterologous to said cell;

wherein said first enzyme is bound to said first binding site and said second enzyme is bound to said second binding site.

2. (Amended) A cell containing a composition comprising:

a) nucleic acid encoding an [exogeneous]exogenous scaffold having no enzymatic activity and comprising at least a first binding site and a second binding site; and

b) nucleic acid encoding at least a first and a second enzyme, wherein at least one of said enzymes is heterologous to said cell;

wherein said first enzyme is capable of being bound to said first binding site and said second enzyme is capable of being bound to said second binding site.

3. A cell according to claim 1 or 2, wherein said scaffold comprises at least three binding sites.

4. A cell according to claim 1 or 2, wherein said scaffold comprises at least four binding sites.

5. A cell according to claim 1 or 2, wherein said scaffold comprises at least five binding sites.

6. A cell according to claim 1 or 2, wherein said binding sites are on the same scaffold molecule.

7. A cell according to claim 1 or 2, wherein said binding sites are on different scaffold molecules.

93/ 8. (Amended) A cell according to claim 1 or 2, further comprising
c) an [exogeneous] exogenous bioactive agent precursor.

5 9. A method of screening for a bioactive agent, said method comprising:

- 10 a) expressing in a plurality of host cells nucleic acid encoding an exogeneous scaffold comprising at least a first binding site and a second binding site;
b) expressing in said plurality of host cells nucleic acids encoding at least a first enzyme and a second enzyme; under conditions where said nucleic acids are expressed, and said first enzyme binds to said first binding site and said second enzyme binds to said second binding site;
c) screening said host cells for a cell exhibiting an altered phenotype, wherein said altered phenotype is due to the presence of a bioactive agent.

15 10. A method of screening for a bioactive agent, said method comprising:

- 15 a) expressing in a plurality of host cells a library of nucleic acids encoding a library of scaffolds, each scaffold comprising at least a first binding site and a second binding site;
b) expressing in said cells a library of nucleic acids encoding a library of enzymes; under conditions where said nucleic acids are expressed, and at least some of said enzymes bind to said scaffolds;
20 c) screening said host cells for an altered phenotype.

11. A method according to claim 9, wherein said expressing step further comprises introducing said nucleic acids into said cells.

12. A method according to claim 10, wherein said introduction comprises retroviral infection.

25 13. A method according to claim 9 further comprising adding at least one exogenous precursor to said cell.

~~27. A cell according to claim 1 or 2, wherein said cell is a mammalian cell.~~

28. A cell according to claim 1 or 2, wherein said scaffold is linear.

~~29. A cell according to claim 1 or 2, wherein said scaffold is circular.~~

3

~~30. A cell according to claim 1 or 2, wherein said scaffold is branched.~~

31. A cell according to claim 1 or 2, wherein said scaffold further comprises a fusion partner.

32. A cell according to claim 1 or 2, wherein at least one of said enzymes further comprises a fusion partner.

~~33. A cell according to claim 31, wherein said fusion partner is a presentation structure.~~

34. A cell according to claim 31, wherein said fusion partner is a targeting sequence.

~~35. A cell according to claim 31, wherein said fusion partner is a rescue sequence.~~

~~36. A cell according to claim 31, wherein said fusion partner is a stability sequence.~~

~~37. A cell according to claim 31, wherein said fusion partner is a linker sequence.~~

~~38. A cell according to claim 32, wherein said fusion partner is a presentation structure.~~

39. A cell according to claim 32, wherein said fusion partner is a targeting sequence.

~~40. A cell according to claim 32 wherein said fusion partner is a rescue sequence.~~

~~41. A cell according to claim 32, wherein said fusion partner is a stability sequence.~~

~~42. A cell according to claim 32, wherein said fusion partner is a linker sequence.--~~

A4
cont